

The effects of interactions between testosterone and ghrelin on mean plasma thyroid hormones concentration in male rats

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ABSTRACT

Ghrelin is an orexigenic peptide synthesized mainly by stomach and hypothalamus. Ghrelin decreases secretion of thyroid hormones. Testosterone is an esteroidogenic hormone that exerts stimulatory effects on Hypothalamus-Pituitary-Thyroid (HPT) axis activity. The aim of the present study was to investigate the effect of interactions between the central injection of ghrelin and testosterone on mean plasma thyroid hormones concentration. Twenty male Wistar rats (200-250 g) were randomly divided into four groups. The groups received saline, 5 nmol ghrelin, 1 µg testosterone and simultaneous injection of 5 nmol ghrelin and 1 µg testosterone in third cerebral ventricle in volumes of 3 µl. Blood samples were collected one day before injection and until 12 hours after that. Mean plasma thyroid hormones concentrations were determined by radio-immunoassay (RIA). The results indicated that testosterone significantly increased the mean plasma concentration of T3 and T4 hormones after injection compared to before injection, whereas ghrelin significantly decreased the mean plasma concentration of T3 and T4 compared to before injection. The results demonstrated that ghrelin significantly decreased the stimulatory effect of testosterone on mean plasma T3 and T4 concentrations.

Keywords: Stimulatory effect, Radio-immunoassay; Hypothalamus-Pituitary-Thyroid.

Introduction

Androgenic-anabolic steroids (AAS) are steroid molecules derived from testosterone (1, 2). AAS Improve athletic ability, balance the energy levels, increase food intake, muscle mass, sexual function, libido, body hair and body weight and decrease risk of osteoporosis in rats (1, 3, 4). Testosterone is physiologically secreted by testes and adrenal glands and transported by the testosterone binding globulins

(TBG) and albumin. TBG is produced in the liver and binds to about two-thirds of serum thyroxine (T4) (5, 6, 7, 8). Peripheral conversion of inactive T4 to biologically active T3 is catalyzed by 5'-deiodinase activity. A recent study showed the specific modulatory effect of testosterone and estradiol on the proliferation of human thyroid carcinoma cell lines, which is independent of Thyrotropin (9). The stimulatory effect of testosterone and estradiol on the expression of TSH mRNA in the pituitary in normal

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Wistar rats is known (10). Ghrelin, a novel 28 amino acid peptide hormone is secreted by the X/A like endocrine cells of stomach, neurons of hypothalamus and some other tissues (11, 12, 13). Ghrelin is involved in a series of biological functions including regulation of energy balance, food intake (14), sleep (15), body weight (16), gastrointestinal motility (17), cardiovascular functions (14) cell proliferation (18), production of pro-inflammatory cytokines (19), and reproduction in many species. Ghrelin is the endogenous ligand for growth hormone secretagogue receptor (GHS-Rs). GHS-R is highly expressed in the ARC with 94% of all NPY-containing neurons co-expressing GHS-R mRNA. The co-localization of the growth hormone secretagogue with these hypothalamic neurons further indicates that the hypothalamus is a main target site for ghrelin (20, 21, 22). During fasting, ghrelin is secreted by stomach cells and neurons of the hypothalamus and following food consumption mean plasma level of ghrelin decreases (23). A recent study in male rats has reported a decreased activity of the HPT axis after ICV injection of ghrelin every 24h for 5 days; pituitary TSH cells were smaller and TSH plasma levels were lower and as a result, thyroid follicles were less active and mean plasma concentration of thyroxine (T4) was reduced compared with saline-treated rats (24). It was shown that injection of 50 μ g acylated ghrelin decreases T3, T4 and TSH from 2 until 7 hour after injection (25). The hypothalamic-pituitary-thyroid (HPT) axis plays an important role in the energy balance via the effect of thyroid hormones to increase oxygen consumption and heat generation. It has been shown that different neural, hormonal and environmental factors interact to modulate thyroid hormones secretions. Hormones of the HPT axis are crucially involved in maintaining body temperature and energy balance in mammals (20, 22).

Earlier studies showed the effect of ghrelin or testosterone on mean plasma thyroid hormones but the simultaneous effect of ghrelin and testosterone on T3 and T4 was unknown. The purpose of this study was to examine the effect of the interaction between ghrelin and testosterone (injected via ICV route) on mean plasma T3 and T4 concentrations in male rats and to assess the potential pathway of this change.

Material and Methods

Animals

Twenty male Wistar rats (200-250 g) used in this study were provided by the Center of Neuroscience Research of Shahid Beheshti University. Animals were housed individually in cages under controlled temperature (25 \pm 2 $^{\circ}$ C) and light (12h light/dark cycle). They had free access to food and water all the time. The rats were divided to four groups and each group included five animals. Group 1 (control) included the animals that received saline (3 μ l ICV), in group 2 (ghrelin) the animals received 5 nmol ghrelin dissolved in saline, in group 3 (testosterone) the animals received 1 μ g testosterone dissolved in sesame oil, and in group 4 (ghrelin+testosterone) the animals received ghrelin and testosterone simultaneously. All injections was done in 3 μ l volumes.

ICV cannulation and injection

Animals were anesthetized by intraperitoneal (IP) injection of a mixture of ketamine and xylazine (100 mg/kg BW ketamine+15 mg/kg BW xylazine). For implanted cannulae, animals were placed in a stereotaxic frame and 22-gauge stainless steel intracerebroventricular (ICV) cannulae were implanted in the third cerebral ventricle 4-6 days before the injection experiments. A cannulae were inserted using the following stereotaxic coordinates: AP-2.3, ML-0.0, DV-6.5 according to coordinates of Paxinos and Watson atlas. The cannulae were fixed to the skull with three stainless steel screws and acrylic dental cement. Dummy cannulae were inserted to prevent the entry of foreign material. One week after surgery, the drugs were injected by 27-gauge stainless steel injector which connected to 5 μ l Hamilton syringe by PE-20 tubing. Ghrelin dose injected was 5 nmol and testosterone dose injected was 1 μ g according to earlier studies (26, 27)

Blood Collection

Blood samples were collected before injection and, 30 minutes, 6 and 12 hours after injection. All injections and blood collections was done between 8-10 am. Correct cannula placement was checked by postmortem dye Natt. Only those animals with properly positioned cannulae were included in data analysis.

Hormones assay

Blood samples centrifuged immediately for 20 min at 4000 rpm and the plasma frozen at -25°C until T3 and T4 concentration were assayed. Plasma T3 and T4 were measured by T3 and T4 kits (Padtangostar, Tehran, Iran) and concentration were determinate by radio-immunoassay (RIA).

Statistical analysis

Data were presented as mean ±SE, and analyzed by repeated measures test followed by post hoc Least Significant Difference and SPSS software. $P < 0.05$ was considered to be statistically significant.

Results

Effect of ghrelin on T3 and T4 concentration

The results showed that ghrelin decreased mean plasma T3 concentration 6 and 12h after injection, respectively, 38.35% and 34.24% compared to before injection (Fig. 1) also decreased mean plasma T4 concentration 30 min, 6 and 12 h after injection, respectively 24.17%, 31.70% and 30.46% compared to before injection (Fig. 2).

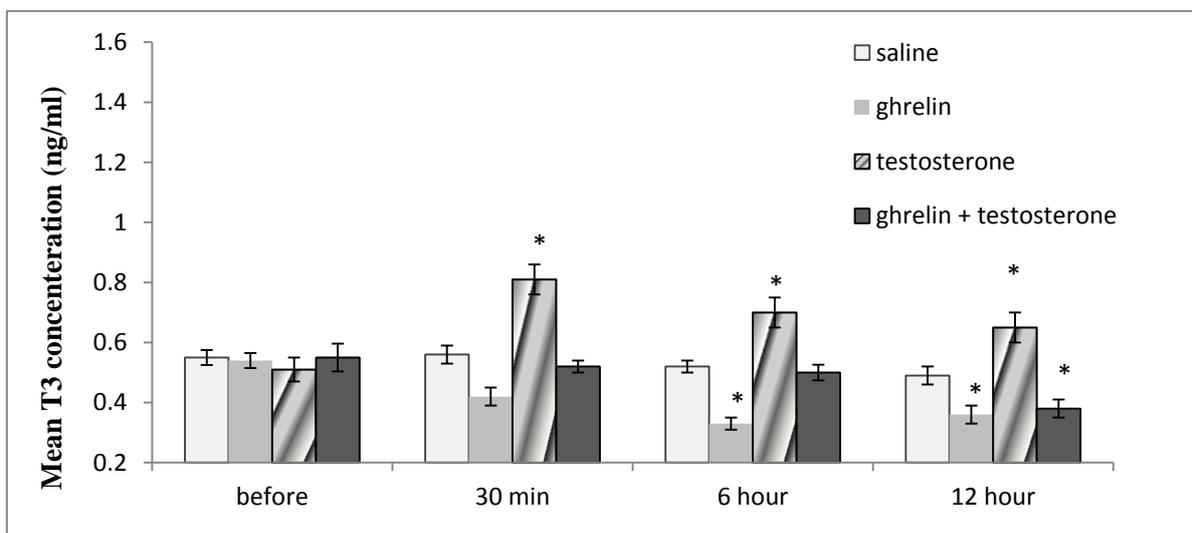


Figure 1. Effect of ghrelin (5 nmol), testosterone (1 µg) and simultaneous injection of ghrelin and testosterone on mean plasma T3 concentration compared to saline in 30 mins, 6 and 12 hours after injection ($p < 0.05$).

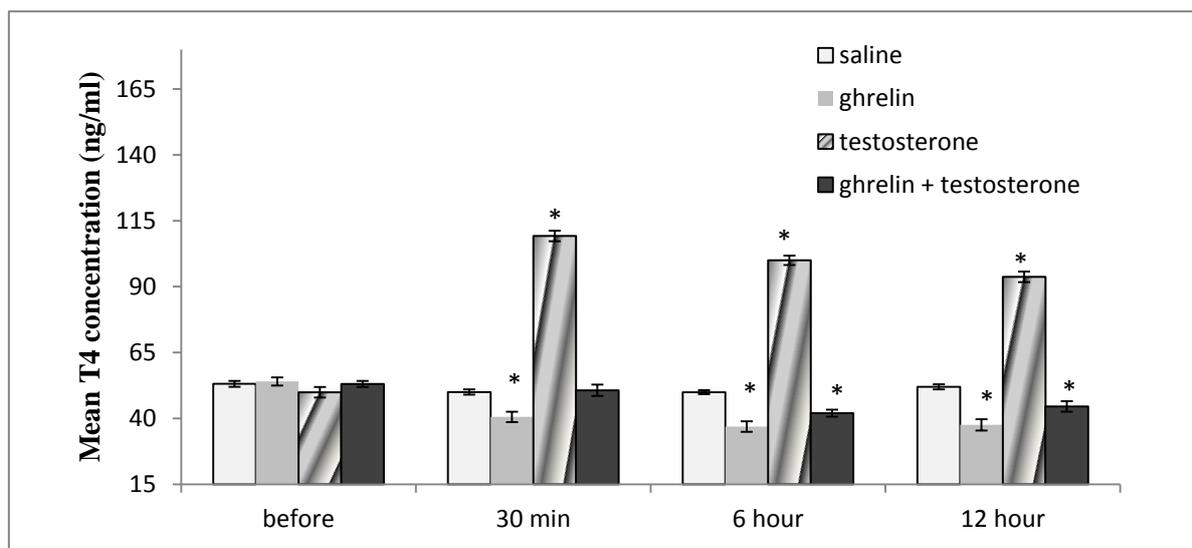


Figure 2. Effect of ghrelin (5 nmol), testosterone (1 µg) and simultaneous injection of ghrelin and testosterone on mean plasma T4 concentration compared to saline in 30 mins, 6 and 12 hours after injection ($p < 0.05$).

Effect of testosterone on T3 and T4 concentration

Testosterone increased mean plasma T3 concentration 30 min, 6 and 12h after injection respectively, 135% , 96.07% and 60.78% compared to before injection ($P<0.05$) (Fig. 1) also increased mean plasma T4 concentration 30 min, 6 and 12h after injection, respectively, 198.91%, 121.61% and 87.70% compared to before injection ($P<0.05$) (Fig. 2).

Effect of ghrelin and testosterone on T3 and T4 concentration

Simultaneous injection of ghrelin and testosterone decreased mean plasma T3 concentration 12h after injection, 30.9% compared to before injection ($P<0.05$) (Fig. 1) also decreased mean plasma T4 concentration, 6 and 12h after injection, respectively, 20.83% and 18.06% compared to before injection ($P<0.05$) (Fig. 2).

Discussion

It seems that significant decrease in T3 and T4 concentration after ghrelin injection was due to effect of ghrelin on different peptides like Agouti-related peptide (AgRP) or neuropeptide Y (NPY) in the ARC of hypothalamus. Previous studies have shown that ghrelin increase the synthesis of AgRP/ NPY in ARC nucleus (26, 27, 28). It was found that chronic central administration of ghrelin increases hypothalamic NPY and AGRP mRNA levels. It has been proven that AgRP/NPY immunoreactive axons densely innervate the thyrotropin-releasing hormone (TRH) neurons in the paraventricular nucleus (PVN) of hypothalamus (26). Other study demonstrated that AgRP and NPY have a similarly potent inhibitory action on the pro-TRH gene expression of hypophysiotropic neurons, indicating that both AGRP and NPY may play a major role in the inhibition of the HPT axis during fasting (29, 30, 31, 32). Earlier observations showed that AgRP and NPY administration *in vivo* decreases TSH levels, while *in vitro* prevents TRH release from hypothalamic explants (29, 30, 33, 34, 35). The effect of NPY in reducing serum TSH is probably due to its impact in increasing dopamine utilization in the

median eminence. Dopamine itself might be inhibitory to TRH release from the median eminence (36) and TSH from anterior pituitary thyrotropes (37). So, ghrelin may have an inhibitory effect on HPT axis activity via increasing AgRP and NPY. Some studies have shown that alpha- melanocyte-stimulating hormone (α -MSH) secreting neurons in ARC densely innervate the TRH neurons in PVN. Therefore, there is a significant increase in TSH and thyroid hormones concentration after ICV or paraventricular injection of α -MSH (29, 38, 39). Hence, we could expect inhibitory effect of ghrelin on HPT axis, at least partially, which might be due to an increase in the Agouti level and its antagonist action on α -MSH receptors. It has been recognized that ICV injection of ghrelin blocked the GABA release from AgRP and NPY neurons in hypothalamus. The inhibition of GABA secretion, cause of increasing corticotrophin releasing hormone (CRH) from hypothalamus by activation of CRF neurons and demonstrated CRH and cortisol exert an inhibitory effect of mean plasma T3 and T4 concentration (40, 41, 42).

The stimulatory effect of testosterone on the expression of TSH mRNA in the pituitary in normal Wistar rats is known (43), in other word testosterone would have increased the responsiveness of thyrocytes to TSH in male rats. Nevertheless, one author has reported that testosterone decreased mean plasma TSH concentration (44). Several studies have reported decreased serum TSH concentration after orchidectomy and reversion of this effect by testosterone replacement (45, 46, 47). Oral testosterone administration decreased TBG, but whether testosterone decreases TBG by reduced synthesis or increased clearance is not recognized (48). Testosterone inhibitory effect on HPA axis (49) decreases CRF which is done by increasing in 5-HT (serotonin precursor) concentration (50, 51). Moreover, 5-HT via 5-HT_{1B}Rs, both inhibits AgRP neuronal activity (49). Further evidence illustrates that the direct 5-HT_{2A/2C} receptor agonist DOI injected into the PVN inhibits the effect of NPY on energy intake and metabolism (52) (Fig. 3).

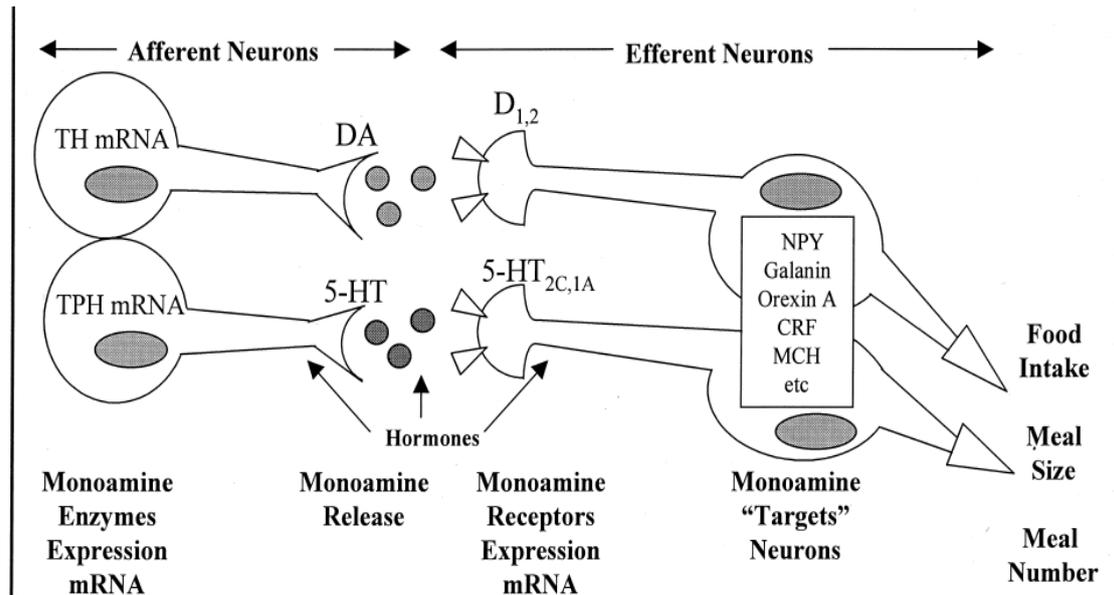


Figure 3. Interaction between NPY, CRF, MCH, 5-HT and dopamine neurons in hypothalamus.

In the present study, the effect of ghrelin+ testosterone on HPT axis was investigated for the first time. We demonstrated that in co-injection of testosterone and ghrelin, ghrelin significantly counteracting effect of testosterone and decreasing mean plasma T3 and T4 concentration.

As mentioned, testosterone decreases CRF by increased 5-HT and eventually 5-HT via 5-HT_{1B}Rs, both inhibit AgRP neuronal activity. Furthermore, we know AGRP plays a major role in the inhibition of the HPT axis. It has also been demonstrated that ghrelin

inhibits 5-HT release from rat hypothalamic (53). So, probably ghrelin decreased the stimulatory effect of testosterone on mean plasma T3 and T4 concentrations by inhibiting 5-HT release and it cause more activity of AgRP neuronal on the TRH neurons in the paraventricular nucleus of hypothalamus.

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REFERENCES

1. Brower, K.J. (2002) Anabolic steroid abuse and dependence. *Curr. Psychiatr. Rep.*, **4**, 377-387.
2. Yesalis, C.E., Kennedy, N.J., Kopstein, A.N., Bahrke, M.S. (1993) Anabolic-androgenic steroid use in the United States. *JAMA*, **270**, 1217-21.
3. Brower, K.J., Blow, F.C., Young, J.P., Hill, E.M. (1991) Symptoms and correlates of anabolic-androgenic steroid dependence. *Brit. J. Addict.*, **86**, 759-768.
4. Dimeo, A.N., Wood, R.I. (2006) ICV testosterone induces FOS in male Syrian hamster brain. *Psychoneuroendocrin.*, **31**, 237-249.
5. Mataradze, G.D., Kurabekova, R.M., Rozen, V.B. (1992) The role of sex steroids in the formation of sex-differentiated concentrations of corticosteroid-binding globulin in rats. *J. Endocrinol.*, **132**, 235-240.
6. Nock, B., Cicero, T.J., Wich, M. (1998) Chronic exposure to morphine decreases physiologically active corticosterone in both male and female rats but by different mechanisms. *J. Pharmacol. Exp. Ther.*, **286**, 875-882.
7. Gala, R.R., Westphal, U. (1965) Corticosteroid-binding globulin in the rat: studies on the sex difference.

Endocrinol., **77**, 841-851.

8. Iqbal, M.J., Dalton, M., Sawers, R.S. (1983) Binding of testosterone and estradiol to sex hormone binding globulin, human serum albumin and other plasma proteins: evidence for non-specific binding of estradiol to sex hormone binding globulin. *Clin. Sci. (Colch.)*, **64**, 307-314.
9. Banu, K.S., Govindarajulu, P., Aruldas, M.M. (2001) Testosterone and estradiol have specific differential modulatory effect on the proliferation of human thyroid papillary and follicular carcinoma cell lines independent of TSH action. *Endocr. Pathol.*, **12**, 315-327.
10. Franklyn, J.A., Wood, D.F., Balfour, N.J., Ramsden, D.B., Docherty, K., Sheppard, M.C. (1987) Modulation by estrogen of thyroid hormones effects on thyrotropin gene expression. *J. Endocr.*, **115**, 53-59.
11. Kojima, M., Hosoda, H., Date, Y., Nakazato, M., Matsuo, H., Kangawa, K. (1999) Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature*, **402**, 656-660.
12. Kojima, M., Kangawa, K., (2005) Ghrelin: structure and function. *Physiol. Rev.*, **85**, 495-522.
13. St Pierre, D.H., Wang, L., Tache, Y. (2003) Ghrelin: a novel player in the gut-brain regulation of growth hormone and energy balance. *News Physiol. Sci.*, **18**, 242-246.
14. Iglesias, M.J., Pineiro, R., Blanco, M. (2004) Growth hormone releasing peptide (ghrelin) is synthesized and secreted by cardiomyocytes. *Cardiovascular Res.*, **62**, 481-488.
15. Seoane, L.M., Al-Massadi, O., Lage, M., Dieguez, C., Casanueva, F.F. (2004) "Ghrelin: from a GH-secretagogue to the regulation of food intake, sleep and anxiety. *Pediat. Endocrinol. Rev.*, **1**, 432-437.
16. Castñeda, T.R., Tong, J., Datta, R., Culler, M., Tschop, M.H. (2010) Ghrelin in the regulation of body weight and metabolism. *Front. Neuroendocrin.*, **31**, 44-60.
17. Peeters, T.L. (2006) Potential of ghrelin as a therapeutic approach for gastrointestinal motility disorders. *Curr. Opin. Pharmacol.*, **6**, 553-558.
18. Granata, R., Settanni, F., Biancone, L., Trovato, L., Nano, R., Bertuzzi, F., Destefanis, S., Annunziata, M., Martinetti, M., Catapano, F., Ghè, C., Isgaard, J., Papotti, M., Ghigo, E., Muccioli, G. (2007) Acylated and unacylated ghrelin promote proliferation and inhibit apoptosis of pancreatic β -cells and human islets: involvement of 3, 5-cyclic adenosine monophosphate/protein kinase A, extracellular signal-regulated kinase 1/2, and phosphatidylinositol 3-kinase/Akt signaling. *Endocrinology*, **148**, 512-529.
19. Taub, D.D. (2007) Novel connections between the neuroendocrine and immune systems: the ghrelin immunoregulatory network. *Vita. Horm.*, **77**, 325-346.
20. Bennett, P.A., Thomas, G.B., Howard, A.D., Feighner, S.D., van der Ploeg, L.H., Smith, R.G., Robinson, I.C. (1997) Hypothalamic growth hormone secretagogue-receptor (GHS-R) expression is regulated by growth hormone in the rat. *Endocrinol.*, **138**, 4552-4557.
21. Howard, A.D., Feighner, S.D., Cully, D.F., Arena, J.P., Liberato, P.A., Rosenblum, C.I., Hamelin, M., Hreniuk, D.L., Palyha, O.C., Anderson, J., Paress, P.S., Diaz, C., Chou, M., Liu, K.K., McKee, K.K., Pong, S.S., Chung, L.Y., Elbrecht, A., Dashkevich, M., Heavens, R., Rigby, M., Sirinathsinghji, D.J., Dean, D.C., Melillo, D.G., Patchett, A.A., Nargund, R., Griffin, P.R., DeMartino, J.A., Gupta, S.K., Schaeffer, J.M., Smith, R.G., Van der Ploeg, L.H. (1996) A receptor in pituitary and hypothalamus that functions in growth hormone release. *Science*, **273**, 974-977.
22. Nogueiras, R., Tovar, S., Mitchell, S.E., Rayner, D.V., Archer, Z.A., Dieguez, C., Williams, L.M. (2004) Regulation of growth hormone secretagogue receptor gene expression in the arcuate nuclei of the rat by leptin and ghrelin. *Diabetes*, **53**, 2552-2558.
23. Tschop, M., Wawarta R., Riepl R.L., Friedrich, S., Bidlingmaier, R.M., Landgraf, R., Folwaczny, C. (2001) Post-prandial decrease of circulating human ghrelin levels. *J. Endocrinol. Invest.*, **24**, 19-21.
24. Susic-Jurjevic, B., Stevanovic, D., Milosevic, V., Sekulic, M., Starcevic, V. (2009) Central ghrelin affects pituitary-thyroid axis: histo-morphological and hormonal study in rats. *Neuroendocrinol.*, **89**, 327-336.
25. Kluge, M., Riedl, S., Uhr, M., Schmidt, D., Zhang, X., Yassouridis, A., Steiger, A. (2010) Ghrelin affects

- the hypothalamus-pituitary-thyroid axis in humans by increasing free thyroxine and decreasing TSH in plasma. *Europ.J. Endocrinol.*, **162**, 1059-1065.
26. Kamegai, J., Tamura, H., Shimizu, T., Sugihara, H., Wakabayashi, I. (2001) Chronic central infusion of Ghrelin increases hypothalamic neuropeptide Y and agouti- related protein mRNA levels and body weight in rats. *Diabetes*, **50**, 2438-2443.
 27. Wren, A.M., Small, C.J., Ward, H.L., Murphy, K.G., Dakin, C.L., Taheri, S., Kennedy, A.R., Roberts, G.H., Morgan, D.G., Ghatei, M.A., Bloom, S.R. (2000) The novel hypothalamic peptide ghrelin stimulates food intake and growth hormone secretion. *Endocrinol.*, **141**, 4325-4328.
 28. Wren, A.M., Small, C.J., Abbott, C.R., Dhillon, W.S., Seal, L.J., Cohen, M.A., Batterham, R.L., Taheri, S., Stanley, S.A., Ghatei, M.A., Bloom, S.R. (2001) Ghrelin causes hyperphagia and obesity in rats. *Diabetes*, **50**, 2540-2547.
 29. Fekete, C., Kelly, J., Mihály, E., Sarkar, S., Rand, W.M., Légrádi, G., Emerson, C.H., Lechan, R.M. (2001) Neuropeptide Y has a central inhibitory action on the hypothalamus- Pituitary- Thyroid axis. *Endocrinol.*, **142**, 2606-2613.
 30. Fekete, C., Kelly, J., Mihály, E., Sarkar, S., Rand, W.M., Légrádi, G., Emerson, C.H., Lechan, R.M. (2002) Agouti- Related Protein (AgRP) has a central inhibitory action on the hypothalamus- Pituitary- Thyroid (HPT) axis; Comparisons between the effect of AgRP and Neuropeptide Y on energy homeostasis and the HPT axis. *Endocrinol.*, **143**, 3846-3853.
 31. Kawano, H., Masuko, S. (2000) Beta-endorphin adrenocorticotrophic hormone and neuropeptide Y-containing projection fibers from the arcuate hypothalamic nucleus make synaptic contacts on to nucleus preopticusmedianus neurons projecting to the paraventricular hypothalamic nucleus in the rat. *Neuro. Sci.*, **98**, 555-565.
 32. Legradi, G., Lechan, R.M. (1999) Agouti-Related protein containing nerve terminals innervate thyrotropin releasing hormone neurons in the hypothalamic paraventricular nucleus. *Endocrinol.*, **140**, 3643-3652.
 33. Billington, C.J., Briggs, J.E., Harker, S., Grace, M., Levine, A.S. (1994) Neuropeptide Y in hypothalamic paraventricular nucleus: a center coordinating energy metabolism. *Amer. J. Physiol.*, **266**, 1765-1770.
 34. Harfstrand, A., Eneroth, P., Agnati, L., Fuxe, K. (1987) Further studies on the effects of central administration of neuropeptide Y on neuroendocrine function in the male rat: relationship to hypothalamic catecholamines. *Regul. Peptides*, **17**, 167-179.
 35. Wittmann, G., Liposits, Z., Lechan, R.M., Fekete, C. (2002) Medullary adrenergic neurons contribute to neuropeptide Yergic innervation of hypophysiotrophic thyrotropin releasing hormone- synthesizing neurons in rats. *Neurosci. Lett.*, **324**, 69-73.
 36. Andersson, K., Eneroth, P. (1987) Thyroidectomy and central catecholamine neurons of the male rat. Evidence for the existence of an inhibitory dopaminergic mechanism in the external layer of the median eminence and for a facilitatory noradrenergic mechanism in the paraventricular hypothalamic nucleus regulating TSH secretion. *Neuroendocrinol.*, **45**, 14-27.
 37. Krulich, L. (1982) Neurotransmitter control of thyrotropin secretion. *Neuroendocrinol.*, **35**, 139-147.
 38. Fekete, C., Legradi, G., Mihaly, E., Huang, Q.H., Tatro, J.B., Rand, W.M., Emerson, C.H., Lechan, R.M. (2000) alpha-Melanocyte-stimulating hormone is contained in nerve terminals innervating thyrotropin-releasing hormone-synthesizing neurons in the hypothalamic paraventricular nucleus and prevents fasting-induced suppression of prothyrotropin-releasing hormone gene expression. *J. Neurosci.*, **20**, 1550-1558.
 39. Sakurazawa, N., Mano-Otagiri, A., Nemoto, T., Shibasaki, T. (2013) Effects of intracerebroventricular ghrelin on food intake and Fos expression in the arcuate nucleus of the hypothalamus in female rats vary with estrous cycle phase. *Neurosci. Lett.*, **541**, 204-208.
 40. Cowley, M.A., Smith, R.G., Diano, S., Tschöp, M., Pronchuk, N., Grove, K.L., Strasburger, C.J., Bidlingmaier, M., Esterman, M., Heiman, M.L., Garcia-Segura, L.M., Nillni, E.A., Mendez, P., Low, M.J., Sotonyi, P., Friedman, J.M., Liu, H., Pinto, S., Colmers, W.F., Cone, R.D., Horvath, T.L. (2001) The

distribution and mechanism of action of ghrelin in the CNS demonstrates a novel hypothalamic circuit regulating energy homeostasis. *Neuroendocrinol.*, **73**, 203-214.

41. Giordano, R., Picu, A., Broglio, F., Bonelli, L., Baldi, M., Berardelli, R., Ghigo, E., Arvat, E. (2004) Ghrelin, hypothalamus- Pituitary- Adrenal axis and cushing's syndrome. *Pituitary*, **7**, 243-248.
42. Jaszberenyi, M., Bujdoso, E., Bagosi, Z., Telegdy, G. (2006) Mediation of the behavioral, endocrine and thermoregulatory actions of ghrelin. *Horm. Behav.*, **50**, 266-73.
43. Fang, X.L., Shu, G., Yu, J.J., Wang, L.N., Yang, J., Zeng, Q.J., Cheng, X., Zhang, Z.Q., Wang, S.B., Gao, P., Zhu, X.T., Xi, Q.Y., Zhang, Y.L., Jiang, Q.Y. (2013) The anorexigenic effect of serotonin is mediated by the generation of NADPH oxidase-dependent ROS. *PLOS ONE*, **8** , 531-542.
44. Akinsanya, K.O., Ghatei, M.A., Bloom, S.R. (1995) Gonadal steroids regulate rat anterior pituitary levels of TSH- releasing hormone and pyroglutamyl-glutamylproline amide-like immunoreactivity. *Endocrinol.*, **136**, 734-740.
45. Pekary, A.E., Knoble, M., Garcia, N.H., Bhasin, S., Hershman , J.M. (1990) Testosterone regulates the secretion of thyrotrophin-releasing hormone (TRH) and TRH precursor in the rat hypothalamic-pituitary axis. *Endocrinol.*, **125**, 263-270.
46. Clarke, I.J., Rao, A., Chilliard, Y., Delavaud, C., Lincoln, G.A. (2003) Photoperiod effects on gene expression for hypothalamic Appetite-regulating peptides and food intake in the ram. *Amer. J. Physiol.*, **284**, 101-115.
47. Farbota, L., Hofmann, C., Oslapas, R., Paloyan, E. (1987) Effects of age and testicular function on the pituitary-thyroid system in male rats Surgery. *J. Endocrinol.*, **102**, 1081-1087.
48. Chetkowski, R.J., Meldrum, D.R., Steingold, K.A., Randle, D., Lu, J.K., Eggena, P., Hershman, J.M., Alkjaersig, N.K., Fletcher, A.P., Judd, H.L. (1986) Biologic effects of transdermal estradiol. *New Engl. J. Med.*, **314**, 1615-1620.
49. Viau, V., Meaney, M.J. (1996) The inhibitory effect of testosterone on hypothalamic-pituitary-adrenal responses to stress is mediated by the medial preoptic area. *The J. Neurosci.*, **76**, 1866-1876.
50. Robichaud, M., Debonnel, G. (2005) Estrogen and testosterone modulate the firing activity of dorsal raphe nucleus serotonergic neurones in both male and female rats. *J. Neuroendocrinol.*, **17**, 179-185.
51. Ross, D.D. (1990) Testosterone increases TSH-beta mRNA and modulates alpha-subunit mRNA differentially in mouse thyrotropic tumor and castrated rat pituitary. *Horm. Metab. Res.*, **22**, 163-169.
52. Currie, P.J. Coscina, D.V. (1997) Stimulation of 5-HT_{2A/2C} receptors within specific hypothalamic nuclei differentially antagonizes NPY-induced feeding. *Neuro. Report.*, **8**, 3759- 3762.
53. Ghersi, M.S., Casas, S.M., Escudero, C., Carlini, V.P., Buteler, F., Cabrera, R.J., Schiöth, H.B., de Barioglio, S.R. (2011) Ghrelin inhibited serotonin release from hippocampal slices. *Peptides*, **32**, 2367-2371.